

Design, Development and Utility of Glycosyl Donors Bearing an Acetoxymethoxy Leaving Group

Hari Babu Mereyala* and Srinivas Reddy Gurralla

Organic Division-III, Indian Institute of Chemical Technology, Hyderabad 500007, India

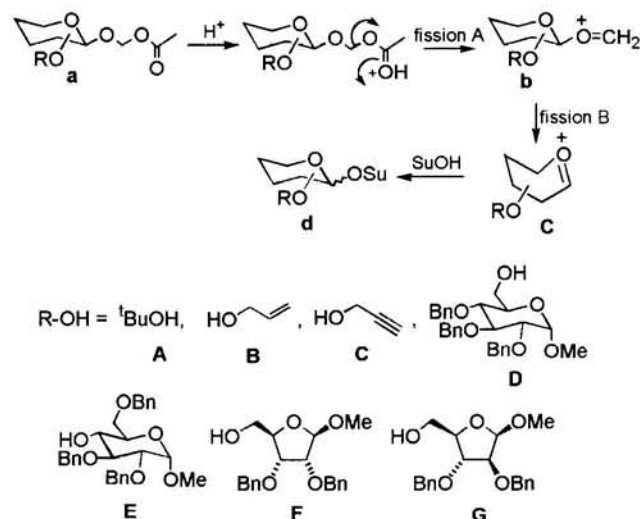
(Received April 27, 1998; CL-980320)

Novel glycosyl donors **2d**, **4d** and **6d** bearing an 'acetoxymethoxy' leaving group have been prepared and their utility is shown by coupling them with alcohols **A-G** to obtain di- and trisaccharides by use of $\text{BF}_3\text{-Et}_2\text{O}$ as a promoter.

A revival in oligosaccharide synthesis has been witnessed¹ due to the increased appreciation of the glycoconjugates for the significant role² played in eukaryotic biology and disease. Highly practical synthesis of complex oligosaccharides involves coupling (inter- and intramolecular³) of a protected glycosyl donor bearing a leaving group at the anomeric centre with a sugar alcohol by use of a suitable activator. Thus, vibrant activity in the designed⁴ and serendipitous⁵ synthesis of glycosyl donors having a variety of anomeric leaving groups (including those generated *in-situ*⁶) has resulted in the development of several glycosylation methods. Contemporary oligosaccharide synthesis relied upon use of glycosyl donors with a leaving group, rather than those without; for the simple reason that they are easy to prepare, have longer shelf life, allow directed activation (armed-disarmed mode⁷) and offer choice of a wide range of activators and solvents to achieve anomeric diastereoselectivity.

Herein, we report our efforts that culminated in the design, development and ultimately utility of a novel protected glycosyl donor having an 'acetoxymethoxy' group that can be activated either by Lewis or protic acids for the synthesis of saccharides. Thus, in principle a protected glycosyl donor **a** bearing an 'acetoxymethoxy' leaving group, when activated by an acid would generate glycosyloxymethylene carbocation **b** (fission A) (Scheme 1), which on further relay (fission B) should lead to the formation of thermodynamically more stable glycosyl cation **c**, to be captured by a nucleophile (sugar alcohol) to form saccharide **d**.

Thus, the glycosyl acetoxymethoxy donor, (2,3,4,6-tetra-O-benzyl- β -D-glucopyranosyloxy)methylene acetate **2d** was synthesized (Scheme 2) by reaction of β -D-glucopyranose pentaacetate **1a** with propargyl alcohol and $\text{BF}_3\text{-Et}_2\text{O}$ in dichloromethane at room temperature for 2 h to obtain the propargyl glycoside **1b**.⁸ Deacetylation (cat. NaOMe/MeOH) followed by benzylation (BnBr/NaH/DMF) of **1b** gave the propargyl derivative **2b**. Reaction of **2b** with $\text{Hg}(\text{OAc})_2$ (0.2 mole equivalent) in 30% aqueous acetone at room temperature for 16 h gave the keto compound **2c**, subsequent Baeyer-Villiger oxidation ($m\text{-CPBA/CH}_2\text{Cl}_2/\text{RT}/6\text{ h}$) gave **2d** in 64.8% overall yield from **1a** as a syrup and was characterized by the $^1\text{H-NMR}$ spectrum from the appearance of a methyl group at δ 1.96 and methylenedioxy protons (2 H) at δ 5.4 (dd). $^{13}\text{C-NMR}$ spectrum indicated C-1 at δ 101.5, an ester carbonyl at δ 170.1 and methylenedioxy carbon at δ 85.4.⁹ IR spectrum showed the ester carbonyl absorption at 1740 cm^{-1} .



Scheme 1.

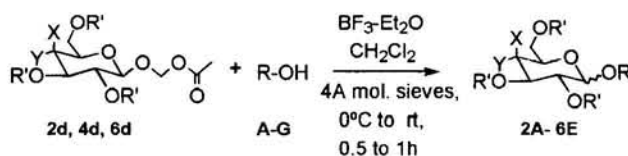
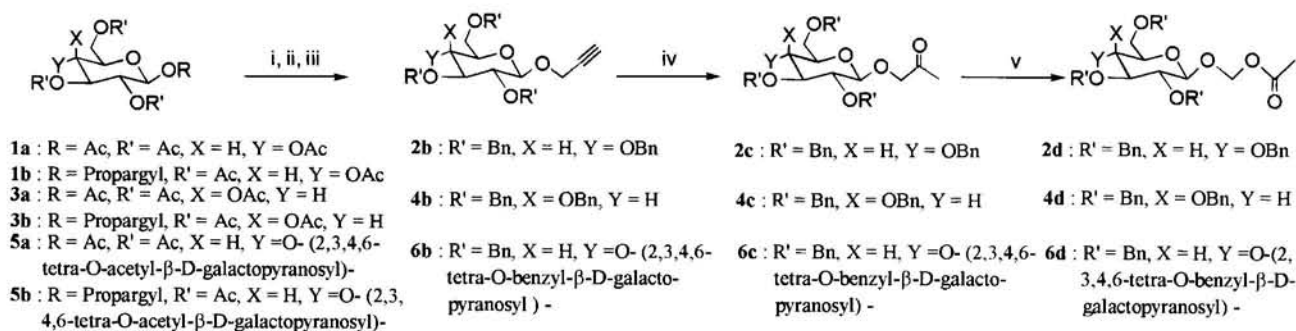


Table 1. Reactions of glycosyl donors **2d**, **4d** and **6d** with various alcohols **R-OH** (**A-G**) to form saccharides **2A-6E**

Entry	Coupling	Saccharides	$\alpha : \beta$ ratio	% yield
1	2d + A	2A	1 : 3	83
2	2d + B	2B	1 : 1	85
3	2d + D	2D	1 : 3	72
4	2d + E	2E	1 : 4	78
5	2d + F	2F	1 : 3	78
6	2d + G	2G	1 : 3	75
7	4d + A	4A	1 : 3	81
8	4d + C	4C	1 : 1	88
9	4d + D	4D	1 : 3	77
10	4d + E	4E	1 : 4	82
11	6d + D	6D	1 : 3	79
12	6d + E	6E	1 : 4	80

Coupling of **2d** with highly reactive **B, D, F, G** and hindered alcohols **A, E** in $\text{CH}_2\text{Cl}_2, \text{BF}_3\text{-Et}_2\text{O}$ at room temperature gave saccharides **2B, 2D, 2F, 2G, 2A** and **2E** respectively in good yields (72-88%) (Table 1), where β -glycosides predominated. Likewise, (2,3,4,6-tetra-O-benzyl- β -D-galactopyranosyloxy)methylene acetate **4d**¹⁰ and the disaccharide donor (2,3,4,6, 2', 3'6'-hepta-



Scheme 2. Reagents and conditions: i: Propargyl alcohol, $\text{BF}_3\text{-Et}_2\text{O}$, CH_2Cl_2 , rt, 2 h, 92-95%; ii: Cat. NaOMe, MeOH, rt, 6 h, iii: NaH, BnBr, DMF, 0°C to rt, 3 h, 90-97%; iv: $\text{Hg}(\text{OCOCF}_3)_2$, 30% aq. acetone, rt, 16 h, 84-91%; v: m-CPBA, CH_2Cl_2 , rt, 6 h, 87-89%.

O-benzyl-β-D-lactopyranosyloxy)methylene acetate **6d**¹¹ were synthesized from the corresponding peracetates **3a** and **5a** respectively in good yields. Coupling of **4d** with alcohols **A**, **C**, **D**, **E** and **6d** with **D** and **E** has resulted in the isolation of saccharides **4A**, **4C**, **4D**, **4E** and **6D**, **6E**¹² respectively in good yields (Table 1).

In conclusion, we have described synthesis of novel glycosyl donors bearing an acetoxymethoxy leaving group and demonstrated its utility for the synthesis of di- and trisaccharides. Efforts are continuing to optimize reaction conditions, such as change of promoter, solvents and temperature to achieve higher diastereoselectivity in the coupling.

Typical experimental procedure: to a mixture of donor (1 mole equiv.) and acceptor (1.1 mole equiv.) in CH_2Cl_2 (3 ml) at 0°C under nitrogen atmosphere was added 4 Å molecular sieves (10 mg) followed by $\text{BF}_3\text{-Et}_2\text{O}$ (1 mole equiv.) and was stirred at RT for 0.5-1 h. When t.l.c. indicated completion of the reaction K_2CO_3 (45 mg) was added, stirred, filtered to remove the solid residue and the filtrate was diluted with CH_2Cl_2 washed with water, organic phase dried (Na_2SO_4) and evaporated to a residue which was purified by column chromatography to obtain the saccharides.

SRG thanks UGC, New Delhi for financial support.

References and Notes

- P. Fugedi, P.J. Garegg, H. Lohn, and T. Norberg, *Glycoconjugate J.*, **4**, 97 (1987); K. Toshima and K. Tatsuta, *Chem. Rev.*, **93**, 1503 (1993); G. J. Boons, *Contemporary Organic Synthesis*, **3**, 173 (1996).
- A. Varki, *Glycobiology*, **3**, 97 (1993); R. A. Dwek, *Chem. Rev.*, **96**, 683 (1996).
- G. Stork, and G. Kim *J. Am. Chem. Soc.*, **114**, 1087 (1992); F. Barresi and O. Hindsgaul, *Can. J. Chem.*, **72**, 1447 (1994); Y. Ito and T. Ogawa, *J. Am. Chem. Soc.*, **119**, 5562 (1997).
- S. Hanessian, C. Bacquet and N. Lehong, *Carbohydr. Res.*, **80**, c17 (1980); R. R. Schmidt and J. Michel, *Angew. Chem., Int. Ed. Engl.*, **19**, 731 (1980).
- D. R. Mootoo, P. Konradsson, U. Udodong and B. Fraser-Reid, *J. Am. Chem. Soc.*, **110**, 5583 (1988).
- B. A. Garcia, J. L. Poole, and D. Y. Gin, *J. Am. Chem. Soc.*, **119**, 7597 (1997); H. Uchiro, K. Miyazaki and T. Mukaiyama, *Chem. Lett.*, **1997**, 403.
- H. B. Mereyala and G. V. Reddy, *Tetrahedron*, **47**, 6435 (1991).
- H. B. Mereyala and S. R. Gurralla, *Carbohydr. Res.*, in press.
- E. Breitmaier, ¹³C NMR spectroscopy methods and application in organic chemistry, 2nd ed (1978).
- Spectral data for **4d**: IR (neat) 1742 cm^{-1} ; $[\alpha]_D -16.5^\circ$ (c, 1.0, CHCl_3); ¹H-NMR (200 MHz, CDCl_3): δ 1.98 (s, 3 H, OCOCH_3), 3.42 - 3.95 (m, 6 H, H_{2-6}), 4.3-5.0 (m, 9 H, H_1 and 4x OCH_2Ph), 5.32, 5.51 (d, 2 H, $J_{\text{gem}} = 7.2\text{ Hz}$, $\text{O-CH}_2\text{-O}$), 7.1-7.4 (m, 20 H, H_{arom}); Selected ¹³C-NMR data (50 MHz, CDCl_3): δ 20.8 (OCOCH_3), 85.3 ($\text{O-CH}_2\text{-O}$), 101.6 (C_1), 170.2 (OCOCH_3).
- Spectral data for **6d**: IR (neat) 1745 cm^{-1} ; $[\alpha]_D 2.06^\circ$ (c, 1.0, CHCl_3); ¹H-NMR (200 MHz, CDCl_3): δ 1.95 (s, 3 H, OCOCH_3), 3.2 - 4.2 (m, 12 H, H_{2-6} , $\text{H}_{2'-6'}$), 4.1 - 5.1 (m, 16 H, $\text{H}_{1,1'}$, 7 x OCH_2Ph), 5.28, 5.5 (d, 2 H, $J_{\text{gem}} = 6.8\text{ Hz}$, $\text{O-CH}_2\text{-O}$), 7.0-7.5 (m, 35 H, H_{arom}); Selected ¹³C-NMR data (50 MHz, CDCl_3): δ 20.8 (OCOCH_3), 85.4 ($\text{O-CH}_2\text{-O}$), 101.5, 102.7 ($\text{C}_{1,1'}$), 170.2 (OCOCH_3).
- All the new compounds gave satisfactory elemental analysis.